Anti-obesity and anti-hyperglycemic effects of ginsenoside Rb1 in rats

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**Objective**— Obesity and type-2 diabetes are national and worldwide epidemics. Since currently available anti-obesity and anti-diabetes drugs have limited efficacy and/or safety concerns, identifying new medicinal agents, such as ginsenoside Rb1 (Rb1) as reported here, offers exciting possibilities for future development of successful anti-obesity and anti-diabetes therapies.

**Research design and methods**—Changes in feeding behavior after acute intraperitoneal (ip) administration of Rb1 and the effects of ip Rb1 for 4 weeks on body weight, energy expenditure, and glucose tolerance in high-fat diet (HFD)-induced obese rats were assessed. We also examined the effects of Rb1 on signaling pathways and neuropeptides in the hypothalamus.

**Results**—Acute ip Rb1 dose-dependently suppressed food intake without eliciting signs of toxicity. This inhibitory effect on feeding may be mediated by central mechanisms because Rb1 stimulated c-Fos expression in brain areas involved in energy homeostasis. Consistent with this, Rb1 activated the PI3K/Akt signaling pathway and inhibited NPY gene expression in the hypothalamus. Four-week administration of Rb1 significantly reduced food intake, body weight gain and body fat content, and increased energy expenditure in HFD-induced obese rats. Rb1 also significantly decreased fasting blood glucose and improved glucose tolerance, and these effects were greater than those observed in pair-fed rats, suggesting that although Rb1’s anti-hyperglycemic effect is partially attributable to reduced food intake and body weight; there may be additional effects of Rb1 on glucose homeostasis.

**Conclusions**— These results identify Rb1 as an anti-obesity and anti-hyperglycemic agent.

Obesity has reached epidemic proportions and created serious public health problems in the United States. Obesity is a major risk factor for type-2 diabetes, cardiovascular problems and some forms of cancer. While efforts to address the environmental and genetic factors responsible for the “epidemic” must continue, and because currently available anti-obesity and anti-diabetic drugs have limited efficacy and/or safety concerns, developing safe and effective medicinal agents, particularly with the dual properties of controlling body weight and reducing blood glucose, offers exciting possibilities for developing successful therapies.

Panax ginseng is a highly valued herb in much of the world, and the root of the ginseng plant has been used as an herbal remedy in Eastern Asia for over 2000 years. The word *panax* is derived from panacea, which means cure-all and longevity. Ginseng and its constituents have historically been ascribed as general tonics and adaptogens to maintain homeostasis and the body’s resistance to adverse factors. The efficacy of ginseng was recognized in the West by the 18th century, and now ginseng is one of the most popular and top-selling herbals in the United States (1:2). Modern pharmacological studies indicate that ginseng extracts possess a wide range of effects, including enhancing learning and memory, reducing anxiety, reducing cancer risk, and improving immunomodulation (3-6).

Chronic administration of an extract of ginseng root significantly reduced body weight, fat mass and serum lipids in diet-induced obese rats (7:8), and extracted ginseng has also been reported to ameliorate...
Rb1’s anti-obese and anti-hyperglycemic effects

hyperglycemia in animals and humans (9-12). In one report, both diabetic and non-diabetic subjects taking an extract of ginseng had stabilized post-prandial glycemia, suggesting that ginseng may also benefit healthy individuals (13). A shortcoming of the existing literature is that the specific compound(s) within extracted ginseng that have beneficial actions are unknown.

Although the components in ginseng root vary across sources and species, ginsenoside Rb1 (Rb1) is considered the most abundant and most important active factor (14). Rb1 has been well characterized chemically (15); it has the structure of a tetracyclic triterpenoid with a molecular formula of C_{54}H_{92}O_{23} and molecular weight of 1109.26 (16). Rb1 has been reported to have diverse biological activities, including facilitating acquisition and retrieval of memory (17), scavenging free radicals (18), blocking calcium over-influx into neurons (19), and preserving the structural integrity of the neurons (20). Rb1 also increased glucose uptake and suppressed lipid accumulation in 3T3-L1 adipocytes, and enhanced insulin secretion from pancreatic Min 6 cells (21-23). Based on these reports, we hypothesized that Rb1 has potent anti-obesity and anti-diabetic effects.

**RESEARCH DESIGN AND METHODS**

**Animals.** Adult male Long-Evans rats (Harlan, Indianapolis, IN) were individually housed in a temperature-controlled vivarium on a 12/12 h light/dark cycle (lights on at 0000 h, lights off at 1200 h). Laboratory chow (Purina 5001, Hudson, NH) was provided ad libitum except where noted. All procedures were approved by the Institutional Animal Care and Use Committee of the University of Cincinnati.

**Materials.** Rb1 purified by high performance liquid chromatography (HPLC) was purchased from Jilin University in China. HPLC (Shimadzu Corp., Kyoto, Japan) analysis was performed in our laboratory and confirmed that the Rb1 had a purity of ≥ 98% using an Rb1 standard obtained from LKT laboratories (St. Paul, MN). The antibody against c-Fos is from Chemicon (Millipore, Billerica, MA), and antibodies for p-Akt and Akt are from Cell Signaling Technology (Beverly, MA).

**Food intake in ad libitum-fed and fasted rats.** Two studies were conducted to determine the effects of acute administration of Rb1 on food intake. In one, ad libitum-fed rats received ip Rb1 in saline at doses of 0, 2.5, 5, 10, or 20 mg/kg; in the second, 24-h fasted rats received ip Rb1 at a dose of 10 mg/kg. The Rb1 or saline was administered at the beginning of dark, and food intake was recorded after 2, 4, 6, 8, 10 and 24 h.

**Conditioned taste aversion assessment.** To determine whether Rb1 elicits visceral illness, we used a standard 2-bottle preference paradigm (24,25). Rats (n = 24) were adapted to 2-h daily access to water (available in two bottles) for 5 days. On Day 6, rats were given 0.15% saccharin in water in each bottle for the 2-h period instead of water and then were injected with either Rb1 (40 mg/kg, ip) or vehicle (n = 8/group). A third group (n = 8) received LiCl as a positive control (0.15 mol/L; 127 mg/kg, ip). On Day 7, animals had water in both bottles during the 2-h session, and on the test day (Day 8), rats were presented with one bottle containing saccharin and one containing water for 24 h. Saccharin preference ratio was calculated as the amount of saccharin consumed divided by the total consumption of both liquids over 2 and 24 h.

**c-Fos expression in the brain.** After a 4-h light-phase food restriction, animals received ip Rb1 (10 mg/kg) or saline at the onset of dark. No food was provided after injection. Rats were anesthetized and perfused with 4% paraformaldehyde for c-Fos immunostaining 2 h after the injections using a previously published method (26). Free-floating sections (30 µm) were cut through the regions of the brainstem and the hypothalamus (27) and
incubated with primary rabbit anti-c-Fos antibody (1:10,000, overnight at 4 °C), biotinylated donkey anti-rabbit antibody (Vector, 1:300, 2 h), and avidin–biotin–peroxidase complex (Vector, 2 h) with PBS wash in between. The sections were then developed by dianinobenzidine with nickel sulfate to produce a dark-purple reaction product. Specific staining was abolished by omission of primary antiserum.

The number of c-Fos positive cells was quantified by a blind observer in the following areas: nucleus of the solitary tract (NTS, -13.80 from bregma) and the hypothalamic arcuate (ARC, -2.30 from bregma), ventromedial (VMH, -2.56 from bregma) and lateral nuclei (LH, -1.88 from bregma). For each animal, the number of c-Fos positive cells was an average value of 4 to 5 sections. Data for c-Fos activation in all of these areas were bilaterally assessed under light microscope with low magnification (×40), and were presented as the total number per section.

**High-fat diet (HFD)-induced obese rats.** Thirty-six rats were fed with a HFD containing 20 g fat (19 g of butter oil and 1 g of soybean oil)/100 g diet prepared by Research Diets, Inc. (Cat. #: D03082706, New Brunswick, NJ) for 13 weeks, as described previously (28). The rats became obese (mean body weight 596 g) and were divided into three groups of comparable body weight. Groups 1 and 2 received ip saline or Rb1 (10 mg/kg), respectively, daily for 28 days and had free access to the same HFD. Because Rb1-treated rats had reduced food intake, a HFD pair-fed group was included as a control. These rats were given the HFD but with an amount limited to the average daily consumption of Rb1-treated rats. Food intake and body weight were recorded daily, energy expenditure and glucose tolerance were assessed at specified times. At the end of the experiment the rats were sacrificed after 6-h fasting and blood and tissues were collected and stored at -80° C for further analyses as detailed below.

**Measurement of intestinal fat absorption.** A non-invasive method was used to measure intestinal fat absorption (29). Feces were isolated from 3 rats in each group on Day 24 of Rb1 treatment, and the fatty acids in the feces were analyzed and quantified with a heptadecanoic acid standard by gas chromatography (29).

**Measurements of plasma samples.** Plasma triglyceride (TG) and cholesterol were measured using Infinity reagent (Thermo, Middletown, VA). Free fatty acid content was measured using NEFA-HR reagent (Wako, Richmond, VA). Leptin was measured using commercially available ELISA kits (Millipore, St. Charles, MO). The levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST) and creatinine were assessed with VITROS Chemistry Products Calibrator Kit 3 and VITROS Chemistry Systems (Ortho-Clinical Diagnostics, Buckinghamshire, UK).

**Indirect calorimetry.** Energy expenditure was assessed with a Physioscan Open Circuit Calorimeter System (Accuscan Instruments, Columbus, OH). The rats were maintained on their respective diet regimens and placed in individual metabolic chambers for 24 h (24). Data were sampled every 60 sec.

**Body fat/lean composition determination.** Body fat was assessed by nuclear magnetic resonance (NMR) (EchoMRI; EchoMedical Systems, Houston, TX). Unanesthetized rats were placed in a restraint tube and inserted into the NMR, providing estimates of total lean tissue, fat tissue and water.

**Histological analysis and morphometry.** Adipose tissue was fixed in 10% formalin for one month and then embedded with paraffin. Tissue sections (10 µm) were cut with a microtome (Leica, Germany) and mounted on superfrost/plus microscope slides. After being air-dried, they were stained with Hematoxylin and Eosin and photographed at
100× magnification. At least two fields per slice and six slices per fat mass were analyzed for the purpose of quantifying adipocyte size. **Intraperitoneal glucose tolerance test (IPGTT).** On Day 28 of treatment, rats were fasted overnight and given ip glucose (1 g/kg, at 1000 h). Blood samples were taken from the tail vein and glucose was assessed with a glucometer (Freestyle, Abbott Diabetes Care Inc. Alameda, CA) at 0 (fasting), 15, 30, 60 120 and 180 min after glucose administration. Plasma insulin was measured in the same samples with a rat insulin ELISA kit (Millipore). **Assessment of the PI3K signaling pathway:** Primary neuronal cells, prepared from hypothalami of embryos and cultured as previously described (30), were treated with different concentrations of Rb1 (0.1-100 µM) for 4 h, or with 10 µM Rb1 for a range of times (0-4 h). The proteins from the cultured neuronal cells, as well as the proteins from the hypothalamus of HFD-induced obese rats with different treatments, were then extracted for measuring phosphorylated Akt (p-Akt) and total Akt levels by Western immunoblotting as described previously (30). **qPCR for neuropeptide mRNA measurement:** Total RNA (100 ng), extracted from hypothalami of HFD-induced obese rats with different treatments using Tri Reagent (Molecular Research Center, Inc., Cincinnati, OH), was reverse-transcribed to first-strand complementary DNA (GE Healthcare Bio-Sciences Corp., Piscataway, NJ). Quantitative real time PCR (qPCR) was performed in a 25-µl final reaction volume with an iCycler iQ Detection System using iQ™ SYBR Green Supermix (Bio-Rad, Laboratories Inc., Hercules, CA). qPCR conditions and sequences of the primers for measuring mRNA levels of the neuropeptides, including proopiomelanocortin (POMC), neuropeptide Y (NPY), and agouti-related protein (AgRP) were described previously (24). **Statistics.** Data were analyzed using parametric statistics (Sigma Stat version 3.1). Two-way ANOVA and two-way repeated-measures ANOVA followed by Student-Newman’s test was used for analysis of food intake, body weight, energy expenditure, blood glucose and plasma insulin levels. For other comparisons, one-way ANOVA was followed by Student-Newman’s test. Data are presented as means ± SE. Values of *P* < 0.05 were considered significant. **RESULTS** **Acute Rb1 administration inhibited food intake.** Ip Rb1 at doses ≥ 5 mg/kg significantly reduced food intake in ad lib-fed rats (Fig. 1A). In a subsequent experiment, ip Rb1 (10 mg/kg) significantly decreased food intake in 24-h fasted rats (Fig. 1B). This effect was present at 4 h lasted for at least 6 h more. There were no significant changes of food intake or body weight by 24 h after Rb1 treatment in any condition (Fig. 1C, D, and E). **Reduction of food intake induced by Rb1 was not associated with illness.** Rats that received saccharin paired with Rb1 (40 mg/kg, ip) had comparable preference for saccharin as saline controls at both 2 and 24 h after treatments. In contrast, animals that received LiCl consumed significantly less saccharin than controls (Fig. 1F). **Rb1 induced brain c-Fos expression.** Rb1 significantly induced c-Fos expression in the hindbrain NTS and the ARC and VMH in the hypothalamus, but not in the LH (Fig. 2). **Rb1 reduced food intake and body weight gain in HFD-induced obese rats.** When Rb1 (10 mg/kg) was administered ip in HFD-induced obese rats, it gradually and significantly decreased food intake during the first week, and the reduced food intake was apparent throughout the 25-day observation period (Fig 3A). Body weight gain was significantly suppressed beginning on Day 8 and continued until the end of the experiment (Fig. 3B). Body weight change was
Rb1’s anti-obese and anti-hyperglycemic effects

comparable between Rb1-treated rats and HFD pair-fed rats during the first 20 days of treatment. However, beginning after the third week, the weight of the Rb1-treated rats became significantly reduced relative to that of the pair-fed rats (Fig. 3B). No side effects, such as diarrhea, were observed. In addition, HFD-fed rats, with or without Rb1, had similar levels of AST, ALT and creatinine (Table 1).

**Rb1 increased energy expenditure in HFD-induced obese rats.** Energy expenditure measured by indirect calorimetry on Day 26 of Rb1 treatment was significantly higher during the dark phase relative to saline controls (Fig. 3C & D). No difference in respiratory quotient occurred among the three groups (data not shown).

**Rb1 reduced body fat in HFD-induced obese rats.** Consistent with the reduced body weight, Rb1-treated rats had reduced body fat compared with saline controls (Fig. 4B vs. 4A). Whereas fat composition was slightly reduced in HFD pair-fed rats, only Rb1-treated rats had a significantly lower percentage of fat compared with controls. There were no differences of lean tissue mass, body water or brown adipose tissue mass among groups (Table 1). To determine whether there was regionally-specific fat loss, selected fat depots were excised and weighed. Rb1 treatment, but not pair-feeding, significantly decreased visceral, including epididymal, retroperitoneal and mesenteric fat, as well as inguinal fat compared to saline controls (Fig. 4C, D, E). Rb1-induced reduction of adipose tissue weight was mainly due to decreased adipocyte size, with adipocytes from epididymal and inguinal fat pads being significantly smaller in Rb1-treated rats compared with saline controls (Fig. 4F and 4G). The reduced body fat was mirrored by a concomitant fall in plasma leptin (Fig. 4H). Rb1 treatment also significantly reduced the levels of plasma TG and cholesterol, but not non-esterified fatty acids (NEFA) (Table 1). There were no differences in these parameters between Rb1-treated and HFD pair-fed rats.

**Rb1 did not affect lipid absorption in HFD-obese rats.** No significant differences in lipid content (%) of the feces were found between Rb1 and saline-treated rats (data not shown), indicating that Rb1 did not affect lipid absorption.

**Rb1 ameliorated hyperglycemia and improved glucose tolerance in HFD-induced obese rats.** After 4 weeks of 10 mg/kg Rb1 treatment, fasting plasma glucose in HFD-induced obese rats was reduced (Fig. 5A), and glucose levels during the IPGTT were reduced by Rb1 all time points from 15-180 min (Fig. 5B). The area under the curve (AUC) in Rb1-treated rats was decreased by 30% compared to vehicle-controls (Fig. 5C). Hence, peripheral Rb1 is efficacious in ameliorating diabetic symptoms associated with obesity. There was also significantly lower fasting insulin in Rb1 treated rats, compared with saline controls (Fig. 5D). Fasting glucose levels and glucose tolerance were improved to a greater extent by Rb1 than by pair-feeding (Fig. 5A and 5C).

**Rb1 activated the PI3K/Akt signaling pathway in hypothalamus.** Phosphorylation of Akt (pAkt) in primary cultured neurons was stimulated by Rb1 at doses of 10 µM or higher (Fig. 6A), with the acute effect of Rb1 on phosphorylation of Akt beginning after 2 h (Fig. 6B). This effect was confirmed in HFD-induced obese rats; i.e., chronic Rb1 treatment, but not pair-feeding, significantly increased hypothalamic pAkt levels (Fig. 6C).

**Rb1 reduced NPY gene expression in HFD-induced obese rats.** Rb1 significantly reduced hypothalamic NPY mRNA (Fig. 6D), but not AgRP mRNA (Fig. 6E), levels compared to saline-controls. While there was a trend toward increased POMC gene expression after Rb1 treatment, it was not significant (Fig. 6F).
DISCUSSION

Ginseng root has historically been ascribed to be a general tonic or adaptogen to maintain the body’s resistance to adverse factors and homeostasis. Modern studies have demonstrated that most of the pharmacological actions of extracted ginseng are attributed to ginsenosides (31), and more than 30 of them have been isolated (32). As a class of compounds, ginsenosides have been demonstrated to be useful in the control of body weight (7;8;11) and blood glucose (9-12;33-36). However, complicated of the multiple known and unknown ingredients in ginseng prevent its application to clinical usage as an alternative medicine to treat obesity and diabetes. The present studies extend previous observations and indicate that ginsenoside Rb1 may prove particularly important in treating obesity and diabetes.

We have demonstrated for the first time that acute ip administration of Rb1 significantly suppresses food intake. This effect is dose-dependent and does not elicit malaise in that an effective dose of Rb1 did not cause a conditioned taste aversion. Hence, visceral illness is likely not a factor in the feeding suppressive actions of peripherally administered Rb1.

c-Fos expression has been used as an index of neuronal activity in rats and mice (37;38). Using c-Fos immunohistochemistry (39), we observed that Rb1, at an effective dose (10 mg/kg) in reducing food intake, significantly induced c-Fos expression in the hindbrain NTS and in the ARC and VMH of the hypothalamus, but not in the LH. Many endogenous factors influence satiation via signals passing from abdominal viscera to the NTS through the vagus nerve, with the signals then being relayed anteriorly to the hypothalamus and other brain areas (40-42). Therefore, one possible explanation of the effect of ip Rb1 on food intake is via alteration of peripheral afferent neural activity. There is also a previous report that food intake was reduced by acute intra-3rd ventricular (i3vt) administration of Rb1 (43), implying that central neural circuits are sensitive to Rb1 and that peripherally administered Rb1 may penetrate the brain reaching local circuits that influence food intake and body weight. Since the blood-brain barrier (BBB) is a major controlling site for the entry of substances into the brain (44;45), evaluating the capability of Rb1 to penetrate the BBB will be an important question for future research. Since ginseng and its extract are generally consumed orally, the implication is that its active ingredients are able to penetrate membranes.

Epidemiological studies have identified a positive correlation between average dietary fat intake and the incidence of obesity. To determine the effect of chronic Rb1 treatment on obesity, we utilized an HFD-induced obese rat model (28). Those studies confirmed Rb1’s effect to reduce food intake, especially during the early period of treatment, and being maintained throughout the observation period, such that Rb1’s anorectic capacity was still evident with sustained administration.

Rb1-treated obese rats behaved normally with no obvious health problems; i.e., no side effects, such as diarrhea were observed during the period of Rb1 treatment. To evaluate whether chronic Rb1 had nonspecific toxic effects that could complicate the interpretation of the results, plasma levels of ALT, AST and creatinine were assessed at the end of the experiment. AST and ALT are enzymes that become elevated with liver disease or injury. Creatinine is a by-product of muscle metabolism and is excreted by the kidneys, and elevated levels can indicate kidney disease, urinary tract obstruction or dehydration. Animals fed HFD, with or without Rb1, had similar levels of AST, ALT and creatinine (Table 1), indicating that chronic treatment with Rb1 did not have any appreciable toxic effect on the liver or kidney.
In parallel with the reduction of food intake, there was a significant decrease in body weight in rats treated with Rb1, compared with vehicle controls. More importantly, the reduced body weight was mainly due to a selective reduction of body fat, leaving lean mass unchanged. Pair-fed rats weighed more than the rats treated with Rb1 after the first three weeks of treatment, implying that Rb1’s effect on body weight is not simply secondary to a reduction of food intake. This is consistent with our observation that chronic treatment with Rb1 significantly increased energy expenditure relative to the saline control group. Although the difference in energy expenditure between Rb1 and pair-fed groups did not reach statistical significance during the one-day test, a slight but persistent increase in energy expenditure induced by Rb1 may explain why Rb1-treated rats had less body weight gain than HFD pair-fed rats beginning after 21 days when food intake was the same for both groups. These observations suggest that attenuated weight gain associated with Rb1 treatment is due to a combination of reduced food intake and increased energy expenditure.

One possible explanation for the reduced fat mass could be reduced fat absorption induced by Rb1. However, this seems unlikely because there was no significant difference in fecal lipid excretion between Rb1 and saline-treated rats, implying that Rb1 did not significantly affect lipid absorption.

Obese subjects are often hyperglycemic and glucose intolerant, and this could be due to increased insulin resistance and/or defects in insulin secretion. Chronic Rb1 treatment significantly reduced fasting glucose and improved impaired glucose tolerance. More importantly, these effects were greater than those observed in pair-fed controls, implying that there are effects of Rb1 on glucose homeostasis beyond those due to decreased food intake and body weight. One way to improve insulin sensitivity is through an increase of tissue glucose uptake, and this possibility is supported by previous in vitro studies demonstrating that Rb1 significantly stimulated basal and insulin-mediated glucose uptake in a time- and dose-dependent manner in both 3T3-L1 adipocytes and C2C12 myotubes (21). The effect was mediated through increased gene expression of GLUT4 and increased GLUT4 translocation to the cell surface (21). Although Rb1-induced reduction of fasting blood glucose was not accompanied by increased plasma insulin in HFD-obese rats, it cannot be ruled out that Rb1 may stimulate insulin secretion because the improved insulin sensitivity requires less insulin to maintain blood glucose level. This possibility is supported by a report that Rb1 promoted glucose-stimulated insulin secretion from Min6 cells, a mouse insulinoma cell line (23).

Phosphatidylinositol-3-kinase (PI3K) is an intracellular signaling pathway that plays a pivotal role in many essential cellular processes. Using primary cultured neuronal cells, prepared from hypothalami of embryos (30), we found that Rb1 significantly and dose-dependently (0.1-100 µM) activates the PI3K signaling pathway beginning after 2 h of treatment. This effect was confirmed in HFD-induced obese rats; i.e., chronic Rb1 treatment, but not pair-feeding, significantly increased hypothalamic pAkt levels, indicating that Rb1 may influence metabolic parameters, at least in part, by regulating this signaling pathway in the brain.

CNS circuits involved in food intake can be conceptually divided into anabolic and catabolic systems. NPY, AgRP and POMC are key peptides in these systems, respectively (46), and it is the balance between these systems that ultimately determines the animal’s ingestive behavior and energy expenditure (46;47). Chronic Rb1 treatment significantly reduced NPY, but not AgRP, mRNA in the hypothalamus of HFD-obese rats, implying that Rb1 exerts its anorectic
action, at least partially, by inhibiting the activity of NPY system. POMC mRNA was non-significantly increased.

The molecular mechanisms as to how Rb1 acts on cells are not clear. Recent reports suggest that Rb1 might act as a phytoestrogen and exert estrogen-like activity through estrogen receptors (16;48). Rb1 activated both ERα and ERβ in dose-dependently, and the activation was inhibited by ICI 182,780, an estrogen receptor antagonist, indicating that these effects were mediated through the ER. Treatment with Rb1 increased ER expression in MCF-7 cells and of AP-1-driven luciferase genes in COS cells (16), but these actions are likely independent of a direct ER association because Rb1 did not displace specific binding of [3H]17β-estradiol from estrogen receptors in MCF-7 whole-cell ligand binding assays (16). These observation suggest that Rb1 is functionally similar to 17β-estradiol. Since estrogen signaling plays an important role in the control of energy homeostasis, whether ER mediates Rb1’s catabolic action will require further study.

In summary, we have demonstrated that ip Rb1 significantly reduces food intake without causing malaise in ad lib-fed or fasted rats. Moreover, chronic ip treatment with Rb1 significantly reduced food intake and body weight gain, and increased energy expenditure in HFD-induced obese rats. Rb1 also significantly decreased fasting blood glucose and improved glucose tolerance to a greater extent than pair-feeding in the HFD-induced obese rats, raising the possibility that Rb1 has important effects on glucose homeostasis. These results identify a novel role for Rb1 as an anti-obesity and anti-hyperglycemic agent.

Author contributions: Y.X. researched data and wrote manuscript; L.S. researched data; K.J.L. researched data; P.T. edited manuscript; Y.Q.X. reviewed manuscript; G.J.W. reviewed manuscript, S.C.W. reviewed/edited manuscript; M.L. wrote and edited manuscript.

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Rb1’s anti-obese and anti-hyperglycemic effects


Table 1. Plasma levels of AST, ALT and creatinine, and body fat and lean composition and lipid profiles in HFD-induced obese rats with or without Rb1 treatment

<table>
<thead>
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<th>Group</th>
<th>Saline</th>
<th>Rb1</th>
<th>Pair-fed</th>
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<tr>
<td>AST (U/L)</td>
<td>119.1±13.7</td>
<td>125.0±10.9</td>
<td>102.8±10.6</td>
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<td>ALT (U/L)</td>
<td>40.0±5.8</td>
<td>37.1±3.6</td>
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<td>Creatinine (mg/dl)</td>
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<td>Fat mass (g)</td>
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<td>140.7±6.2*</td>
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<tr>
<td>Lean mass (g)</td>
<td>377.6±9.3</td>
<td>382.9±4.9</td>
<td>379.2±7.6</td>
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<tr>
<td>Brown adipose tissue (g)</td>
<td>0.94±0.07</td>
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<td>Water (g)</td>
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<td>TG (mg/dL)</td>
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<td>NEFA (mEq/L)</td>
<td>0.442±0.035</td>
<td>0.439±0.023</td>
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*P < 0.05, **P < 0.01, compared with HFD obese rats treated with saline.

**Figure legends**

FIG.1. Ip administration of Rb1 dose-dependently reduced food intake in *ad libitum*-fed rats (A). In 24 h–fasted rats, Rb1 (10 mg/kg, ip) significantly reduced food intake (B). No significant difference in food intake (C, D) or body weight change (E) were found in either ad lib-fed or fasted rats after Rb1 treatment. Ip administered LiCl, but not Rb1 (40 mg/kg, ip), caused a CTA in rats (F). Values are expressed as means ± SE, n = 6 - 8. *P < 0.05, **P < 0.01, compared with saline controls.

FIG. 2. Rb1 (10 mg/kg, ip) significantly increased c-Fos expression in the cells of NTS, VMH, and ARC, but not LH, areas. A-H: Images of representative brain microsections. I: Fos-like immunoreactivity counts (Mean ± SE) in difference brain sections, n = 4-5. *P < 0.05, **P < 0.01, compared with saline controls. cc: central canal; 3V: 3rd ventricle; opt: optic tract.

FIG. 3. Chronic treatment of Rb1 in HFD-induced obese rats significantly reduced food intake (A), body weight gain (B), and increased energy expenditure (C), especially during the dark phase (D), when compared with saline administration. Food intake after Rb1 treatment was
Rb1’s anti-obese and anti-hyperglycemic effects

significantly reduced compared with that after saline treatment since Day 2, and the changes of body weight in Rb1-treated rats were significantly lower than that in saline controls since Day 8. Rb1-treated rats also had significantly less body weight gain vs. rats under pair-feeding since Day 21. In addition, caloric restriction also has elevated energy expenditure, but the difference between saline and pair-fed group is not as prominent as saline and Rb1 group. Mean ± SE, n = 12. *P < 0.05, **P < 0.01, compared with saline controls; dP < 0.05, Rb1-treated vs. pair-fed rats.

FIG. 4. Effect of chronic administration of Rb1 on body fat and plasma leptin levels in HFD-induced obese rats. Whole body weight (A and B), visceral fat tissues, including epididymal, retroperitoneal and mesenteric fat, as well as inguinal fat were significantly decreased after Rb1 treatment (C). D and E are photos of epididymal and mesenteric fat in rats treated with Rb1 or saline. F: H&E staining of epididymal and inguinal fat tissues. The size of adipocytes was calculated and represented by total adipocyte area (n = 6). Rb1 treatment resulted in reduced average adipocyte size (G). Rb1 decreased plasma leptin (H). *P < 0.05; **P < 0.01 vs. saline controls.

FIG. 5. Rb1 reduced fasting blood glucose and improved glucose tolerance in HFD-induced obese rats. (A): Basal hyperglycemia was significantly reduced in Rb1-treated obese rats, compared with both saline controls and pair-fed. (B): After ip administration of glucose (1 g/kg), the glucose incremental area under the curve (AUC) was significantly lower in the Rb1 than the saline group. (C) and (D): Rb1 treatment also reduced fasting insulin level and the insulin AUC after ip glucose. Mean ± SE, n = 12. *P < 0.05; **P < 0.01 vs. saline group; #P < 0.05 vs. pair-fed group.

Fig. 6. Effects of Rb1 on phosphorylation of Akt in cultured primary hypothalamic neurons and in the hypothalamus of HFD-induced obese rats. A: cells were treated with different concentrations of Rb1; B: cells were treated with Rb1 (10 µM) for of the indicated times. n = 3-4. C: Hypothalamic tissues were dissected from HFD-induced obese rats. D: Effect of chronic Rb1 treatment on neuropeptide gene expression in HFD-induced obese rats. n = 6 - 7, mean ± SE, *P < 0.05, compared with before treatment (in vitro) or saline controls (in vivo), and dP < 0.05, compared with pair-fed rats.
Figure 1

A

B

C

D

E

F

Rb1’s anti-obese and anti-hyperglycemic effects
Rb1’s anti-obese and anti-hyperglycemic effects

Figure 2
Figure 4

Figure 5
Figure 6

A) Graph showing the ratio of pAkt/Akt in response to different concentrations of Rb1 (μM).
B) Graph depicting the ratio of pAkt/Akt over time after Rb1 treatment (h).
C) Graph illustrating the ratio of pAkt/Akt for Saline, Rb1, and Pair-fed groups.
D) Bar chart comparing % NPY mRNA in saline-treated rats.
E) Bar chart comparing % AgRP mRNA in saline-treated rats.
F) Bar chart comparing % POMC mRNA in saline-treated rats.