

Genetic Variant in the IGF2BP2 Gene May Interact With Fetal Malnutrition to Affect Glucose Metabolism

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OBJECTIVE—Fetal malnutrition may predispose to type 2 diabetes through gene programming and developmental changes. Previous studies showed that these effects may be modulated by genetic variation. Genome-wide association studies discovered and replicated a number of type 2 diabetes-associated genes. We investigated the effects of such well-studied polymorphisms and their interactions with fetal malnutrition on type 2 diabetes risk and related phenotypes in the Dutch Famine Birth Cohort.

RESEARCH DESIGN AND METHODS—The rs7754840 (*CDKAL1*), rs10811661 (*CDKN2AB*), rs1111875 (*HHEX*), rs4402960 (*IGF2BP2*), rs5219 (*KCNJ11*), rs13266634 (*SLC30A8*), and rs7903146 (*TCF7L2*) polymorphisms were genotyped in 772 participants of the Dutch Famine Birth Cohort Study ($n = 328$ exposed, $n = 444$ unexposed). Logistic and linear regression models served to analyze their interactions with prenatal exposure to famine on type 2 diabetes, impaired glucose tolerance (IGT), and area under the curves (AUCs) for glucose and insulin during oral glucose tolerance testing (OGTT).

RESULTS—In the total population, the *TCF7L2* and *IGF2BP2* variants most strongly associated with increased risk for type 2 diabetes/IGT and increased AUC for glucose, while the *CDKAL1* polymorphism associated with decreased AUC for insulin. The *IGF2BP2* polymorphism showed an interaction with prenatal exposure to famine on AUC for glucose ($\beta = -9.2$ [95% CI -16.2 to -2.1], $P = 0.009$).

CONCLUSIONS—The *IGF2BP2* variant showed a nominal interaction with exposure to famine in utero, decreasing OGTT AUCs for glucose. This may provide a clue that modulation of the consequences of fetal environment depends on an individual's genetic background. *Diabetes* 58:1440–1444, 2009

The fetal origins hypothesis (1,2) states that malnutrition during fetal development predisposes to adverse health outcomes, such as type 2 diabetes. According to the hypothesis, fetal adaptation to a low-caloric intrauterine environment may involve programming to optimize the use of restricted

nutrient supply. Programming may lead to altered gene expression profiles and eventually to disease later in life. In addition, adverse intrauterine circumstances may divert nutrients to critical organs such as the brain at the expense of organs such as the pancreas, liver, or muscles (3).

Type 2 diabetes and defective insulin secretion show high heritabilities (4). Genetic variation is likely to interact with the fetal response to an extreme nutritional situation. We found that the effects of the Pro12Ala polymorphism of the peroxisome proliferator-activated receptor (PPAR) γ 2 gene depend on prenatal exposure to famine (5), and several studies (6–8) have shown interactions of genes with size at birth, a marker of fetal environment. Such interactions may have consequences for development and function. Previously, the *PPAR γ* , *TCF7L2*, and *KCNJ11* genes were identified by linkage and candidate gene studies as type 2 diabetes risk loci (9–11). Recently, genome-wide association studies have identified and replicated new genetic variants associated with type 2 diabetes (12–15). Notably, a number of these variants are thought to be involved in the development and function of critical organs for glucose metabolism, such as the pancreatic β -cell (16–18).

Based on the findings described above, we hypothesized that these genes interact with fetal exposure to famine. In the Dutch Famine Birth Cohort, we investigated the effects of genetic variation in these loci on type 2 diabetes, impaired glucose tolerance (IGT), oral glucose tolerance testing (OGTT), and their interaction with fetal malnutrition.

RESEARCH DESIGN AND METHODS

The Dutch Famine Birth Cohort is composed of individuals born as term singletons around the time of the Dutch famine during World War II in the Wilhelmina Gasthuis in Amsterdam. Details of the study have been described elsewhere (19). In brief, a total of 2,414 singletons were born between 1 November 1943 and 28 February 1947. Members of the cohort living in the Netherlands at 1 September 2002 were invited ($n = 1,423$). Of these, 810 agreed to participate. The study was approved by the local medical ethics committee and was conducted in accordance with the declaration of Helsinki. All participants gave their written informed consent.

Exposure to famine. Prenatal exposure to famine was defined as a daily food ration of the mother $<1,000$ calories during any 13-week period of gestation, based on the official daily food rations for the general population aged ≥ 21 years. Based on these data, individuals born between 7 January 1945 and 8 December 1945 were exposed to famine in utero. Initially, we defined three exposure groups of 16 weeks, dividing the period of exposure in late, mid-, and early gestation. However, this led to small groups in which meaningful genetic analyses were impossible. Therefore, we combined the exposed groups for the interaction analyses.

Study parameters. Trained research nurses conducted measurements and interviews as previously described (19). In brief, information on medical history, lifestyle, and medication was derived from a standardized interview. Information on the mother, the pregnancy, and size at birth was derived from medical birth records.

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See accompanying original article, p. 1428, and commentary, p. 1255.

TABLE 1
General and maternal characteristics according to time of pregnancy when exposed to the Dutch famine

	All	Born before	Late gestation	Mid-gestation	Early gestation	Conceived after	<i>P</i> *
<i>n</i>	772	233	140	117	71	211	
Characteristics at age 58 years							
% men	45.9	46.8	44.3	39.3	42.3	50.7	NS
Age (years)	58.3 ± 0.9	59.2	58.5	58.2	58.0	57.4	NS
BMI (kg/m ²)	28.6 ± 4.8	28.7	28.3	28.1	28.0	29.0	NS
Waist circumference (cm)	92.7 ± 8.9	93.3	92.9	90.8	91.9	93.2	NS
Currently smoking (%)	24.1	20.6	27.9	25.9	31.0	23.1	NS
Birth characteristics							
Gestational age (days)	285 ± 11.1	284	283	285	289	285	NS
Birth weight (g)	3,359 ± 469	3,396	3,187	3,204	3,500	3,471	<0.001
Maternal characteristics							
Age at delivery (years)	28.9 ± 6.4	28.7	31.1	28.8	27.2	28.4	NS
Primiparous (%)	33.7	36.1	20.0	33.3	42.3	37.4	0.04
Weight gain (third trimester)	2.9 ± 2.9	2.8	0.0	4.3	5.0	3.4	0.01
Weight at last antenatal visit	66.4 ± 8.7	66.4	62.8	63.6	69.0	69.3	<0.001

Data are means ± SD. **P* value for comparison between exposed and nonexposed. NS, not significant.

The present study was limited to measures of glucose metabolism. Pre-existent diabetes was defined as the use of oral or injected antidiabetes medications. Individuals with pre-existent diabetes were excluded from OGTT. OGTT was performed after an overnight fast with a standard load of 75 g of glucose at *t* = 0. Blood samples were collected after 0, 30, 60, and 120 min for measurement of plasma glucose and insulin concentrations. Plasma glucose was measured by a standardized enzymatic photometric assay on a Modulator P analyzer (Roche, Basel, Switzerland). Plasma insulin was measured by immunoluminometric assay on an Immulite 2000 Analyzer (Diagnostic Product Corporation, Los Angeles, CA). IGT was defined as a 120-min glucose level between 7.8 and 11.0 mmol/l. Diabetes based on OGTT was defined as a 120-min glucose level >11.0 mmol/l.

Genotyping. Genomic DNA was extracted from fasting blood samples. The rs7754840 (*CDKAL1*), rs10811661 (*CDKN2AB*), rs1111875 (*HHEX*), rs4402960 (*IGF2BP2*), rs5219 (*KCNJ11*), rs13266634 (*SLC30A8*), and rs7903146 (*TCF7L2*) polymorphisms were genotyped with Taqman allelic discrimination assays. The assays were designed and optimized by Applied Biosystems (Foster City, CA). The analyses were performed as described previously (20). Assays were run on 90 blood bank samples to test for adequate cluster separation. Genotypes were determined in 2 ng genomic DNA. Reactions were performed on the Taqman Prism 7900HT platform. Success rates for genotyping ranged from 86.1% (*IGF2BP2*) to 96.7%. The duplicate mismatch rate for the *IGF2BP2* assay was 0. Genotyping for at least one polymorphism was successful in 772 subjects.

Statistical methods. The primary end point was the area under the curve (AUC) for glucose and insulin during OGTT. A secondary combined end point of type 2 diabetes and IGT consisted of pre-existent type 2 diabetes and type 2 diabetes and IGT identified at OGTT.

Continuous variables are expressed as means ± SD. Comparisons between groups were performed with ANOVA and χ^2 tests for normally distributed continuous and categorical variables, respectively. Logarithmic transformation was performed on variables that were not normally distributed. Allele frequencies were estimated by gene counting. Hardy-Weinberg equilibrium was tested by χ^2 testing.

The AUCs during OGTT were calculated by the trapezoidal rule (21): $\{[15 \times \log(\text{gluc0min})] + [30 \times \log(\text{gluc30min})] + [45 \times \log(\text{gluc60min})] + [30 \times \log(\text{gluc120min})]/120\} + \{[15 \times \log(\text{ins0min})] + [30 \times \log(\text{ins30min})] + [45 \times \log(\text{ins60min})] + [30 \times \log(\text{ins120min})]/120\}$ for glucose and insulin, respectively. Binary logistic and linear regression models served to investigate the associations of the genetic variants with type 2 diabetes/IGT risk and AUC for glucose and insulin. The additive model of inheritance was assumed.

Genotype by prenatal famine exposure interactions were tested by creating interaction terms for each genetic variant (coded 0, 1, or 2 for carrying the risk allele) with the exposure group (coded 0 and 1 for unexposed and exposed subjects, respectively). Unexposed subjects were the subjects born before or conceived after the Dutch famine. Genotype by birth weight interaction models were created by taking the product of birth-weight (continuous) and the genetic variant (coded 0, 1, and 2). All models were adjusted for sex. Subsequently, models were adjusted for BMI. Additionally, waist circumference and the mother's parity and weight at the last antenatal visit

were added. Models investigating famine were additionally adjusted for birth weight and vice versa. We adjusted for multiple testing by means of Bonferroni adjustment. We had 80% power to detect interactions of $\beta = 7.8$ with gene and environmental $\beta = 3.0$ (minor allele frequency 30%, $\alpha = 0.05$).

RESULTS

General characteristics and population genetics. General and maternal characteristics of 772 genotyped individuals divided into the famine exposure groups are shown in Table 1. Minor allele frequencies were 0.31, 0.19, 0.35, 0.29, 0.36, 0.29, and 0.30 for the *CDKAL1*, *CDKN2A/B*, *HHEX*, *IGF2BP2*, *KCNJ11*, *SLC30A8*, and *TCF7L2* polymorphisms, respectively. All polymorphisms were in Hardy-Weinberg equilibrium ($\chi^2 < 3.3$; 2 df; *P* > 0.07 for all SNPs). A total of 94 individuals with type 2 diabetes (62 prevalent and 32 based on OGTT) and 100 individuals with IGT were included in the analyses on type 2 diabetes/IGT. For the analyses on the AUCs for glucose and insulin during OGTT, 102 individuals were excluded from the total population because of pre-existent diabetes, nonfasting before the test, or incomplete results.

Associations and interactions with prenatal exposure to famine on type 2 diabetes/IGT and AUCs for glucose and insulin. Table 2 shows odds ratios (ORs) of the genetic variants for the composite outcome type 2 diabetes/IGT and the β -coefficients for AUCs for glucose and insulin during the OGTT. The *TCF7L2* and *IGF2BP2* variants were associated with increased type 2 diabetes/IGT risk (*TCF7L2*: OR 1.39 [95% CI 1.08–1.79], *IGF2BP2*: 1.43 [1.11–1.85]) and increased AUC for glucose (*TCF7L2*: $\beta = 4.5$ [1.0–8.1], *IGF2BP2*: $\beta = 3.6$ [0.1–7.1]). The *CDKAL1* variant associated with a decreased AUC for insulin ($\beta = -8.2$ [-16.1 to -0.41]), which became less strong after adjustment for BMI. None of the other results changed considerably after additional adjustments in multivariate models. None of the polymorphisms had a significant effect on birth weight or were related to prenatal famine exposure (data not shown).

Table 3 shows the interactions between genetic variants and exposure to famine in utero. The *IGF2BP2* showed a significant interaction on AUC glucose (β interaction -9.2 [-16.2 to -2.1], *P* = 0.009). Figure 1 shows the effects of the *IGF2BP2* polymorphism on AUC

for glucose in exposed and unexposed subjects. After Bonferroni correction for 21 tests, this was no longer significant. None of the polymorphisms showed a significant interaction with birth weight (data not shown). Additional adjustments in multivariate models did not change the results.

DISCUSSION

In the present study, we investigated interactions of type 2 diabetes-associated polymorphisms in a population exposed to famine in utero. The *IGF2BP2* polymorphism showed a nominally significant interaction with prenatal exposure to famine on AUC for glucose during OGTT.

The Dutch Famine Birth Cohort provides a unique opportunity to directly investigate the interactions of fetal malnutrition with type 2 diabetes and related measurements. Most studies on the association of early nutrition with type 2 diabetes and related parameters have used birth weight as a marker of fetal nutrition and growth. Unfortunately, a consequent limitation is the size of our study population, which limits the power to detect small effects. This especially holds for the categorical outcome type 2 diabetes/IGT. Nonetheless, effects found in the population largely corresponded to previous literature, except for the direction of the effect of the *HHEX* polymorphism, for which we have no explanation. This effect disappeared after adjustment for BMI. Due to power limitations, we were unable to investigate the interactions with timing of exposure to famine during gestation (late, mid-, or early gestational exposure to famine).

Several studies have identified interactions of size at birth and genetic variants on type 2 diabetes risk and measurements of glucose metabolism (6–8). Previously, we reported an interaction between a *PPARγ* gene variant and fetal malnutrition (5). Now, we observed a nominally significant interaction of the *IGF2BP2* polymorphisms with exposure to famine on glucose levels during OGTT. The effect on type 2 diabetes/IGT risk was in the same direction but not significant. The presence of both the *IGF2BP2* risk allele and exposure to famine associated with lower AUC for glucose, which seems counterintuitive considering the main effects of the two factors, which are both associated with increased type 2 diabetes risk. However, a similar interaction was observed in two other studies on the effect of a genetic variant in the *ACE* gene (6,7). Subjects carrying the *ACE* risk allele were relatively protected from the effect of low birth weight on insulin resistance (6,7). These findings together with the current finding on *IGF2BP2* may be explained by over-efficient modulating effects of risk alleles during fetal development. This would imply that the *IGF2BP2* variant, being part of an important developmental pathway, confers a relative resistance to the detrimental consequences of fetal malnutrition on glucose tolerance at an adult age.

IGF2BP2, also referred to as IMP-2, is an mRNA-binding protein that posttranslationally regulates IGF2, a fetal growth factor, during several developmental stages (22). IGF2 plays a critical role during placental and fetal development (23,24). IGFBPs have a tissue-specific expression pattern, with *IGF2BP2* being expressed in fetal lung, kidney, thymus, and placenta and having its highest expression in fetal liver (22). Interactions between genetic variation in *IGF2BP2* and fetal malnutrition may be exerted through this IGF2 developmental pathway.

TABLE 2
Associations with DM/IGT and AUC for glucose and insulin in the total population

Gene variant	Major/minor allele	OR1 diabetes/IGT (95% CI)		OR2 diabetes/IGT (95% CI)		β1 AUC glucose		β2 AUC glucose		β1 AUC insulin		β2 AUC insulin	
		OR1	(95% CI)	OR2	(95% CI)	β1	(95% CI)	β2	(95% CI)	β1	(95% CI)	β2	(95% CI)
CDKAL1 rs7754840	G/C	1.11	(0.87–1.42)	1.16	(0.90–1.49)	0.04	(–3.3 to 3.4)	0.7	(–2.6 to 4.0)	–8.2	(–16.1 to –0.41)	–6.0	(–13.3 to 1.3)
CDKN2A/B rs10811661	T/C	1.08	(0.81–1.44)	1.03	(0.76–1.39)	–0.004	(–4.0 to 4.0)	–0.4	(–4.2 to 3.5)	–0.22	(–9.4 to 8.9)	–1.4	(–9.9 to 7.1)
HHEX rs1111875	C/T	1.09	(0.86–1.36)	0.99	(0.79–1.27)	–2.1	(–5.2 to 1.0)	–2.7	(–5.8 to 0.3)	2.43	(–4.8 to 9.62)	0.1	(–6.7 to 6.8)
IGF2BP2 rs4402960	G/T	1.43	(1.11–1.85)	1.42	(1.09–1.84)	3.6	(0.1–7.1)	3.2	(–0.2 to 6.5)	3.16	(–4.9 to 11.2)	1.66	(–5.9 to 9.2)
KCNJ11 rs5219	AG (341)	0.97	(0.76–1.23)	0.99	(0.77–1.27)	–1.4	(–4.7 to 1.8)	–1.4	(–4.5 to 1.8)	2.46	(–5.1 to 10.0)	2.9	(–4.1 to 9.9)
SLC30A8 rs13266634	C/T (299)	0.88	(0.68–1.13)	0.91	(0.70–1.18)	0.3	(–3.1 to 3.8)	0.6	(–2.7 to 3.9)	4.26	(–3.7 to 12.2)	4.8	(–2.6 to 12.1)
TCF7L2 rs7903146	C/T (316)	1.39	(1.08–1.79)	1.37	(1.06–1.78)	4.5	(1.0–8.1)	4.6	(1.2–8.0)	–5.5	(–13.5 to 2.52)	–5.3	(–12.9 to 2.1)

OR1 and β1: adjusted for sex. OR2 and β2: adjusted for sex and BMI.

TABLE 3
Interactions of genetic variants with prenatal exposure to famine

Gene variant	OR interaction DM/IGT (95% CI)*	P for interaction	β interaction AUC glucose (95% CI)*	P for interaction	β interaction AUC insulin(95% CI)*	P for interaction
<i>CDKAL1</i> -rs7754840	1.16 (0.70–1.90)	0.57	2.20 (–4.7 to 9.1)	0.66	7.7 (–8.2 to 23.7)	0.37
<i>CDKN2A/B</i> -rs10811661	1.37 (0.75–2.51)	0.31	3.46 (–4.8 to 11.7)	0.42	11.6 (–7.4 to 30.6)	0.24
<i>HHEX</i> -rs1111875	1.61 (1.01–2.58)	0.04	5.2 (–1.2 to 11.5)	0.08	–4.7 (–19.4 to 10.0)	0.55
<i>IGF2BP2</i> -rs4402960	0.82 (0.49–1.37)	0.45	–9.2 (–16.2 to –2.1)	0.009	0.49 (–15.9 to 16.8)	0.94
<i>KCNJ11</i> -rs5219	1.22 (0.75–1.98)	0.41	–2.9 (–9.7 to 3.9)	0.42	–6.0 (–21.6 to 9.7)	0.42
<i>SLC30A8</i> -rs13266634	1.38 (0.82–2.32)	0.22	2.0 (–5.0 to 8.9)	0.62	–7.1 (–23.0 to 8.9)	0.40
<i>TCF7L2</i> -rs7903146	1.12 (0.68–1.85)	0.66	–0.1 (–7.2 to 7.1)	0.93	–0.8 (–15.5 to 17.0)	0.86

*OR and β for the interaction term: All models were adjusted for sex. P values for AUC for glucose and insulin calculated after logarithmic transformation. DM, type 2 diabetes.

However, such explanations are fully speculative, and it is important to stress that our observation was no longer significant after correcting for multiple testing and should therefore be interpreted with caution. Nonetheless, it may suggest that consequences of the fetal environment depend on an individual's genetic background. Interactions of other variants investigated may unfortunately not have become apparent due to the limited power of our study.

None of the polymorphisms was associated with birth weight. This is not in line with the fetal insulin hypothesis, which proposes that the same genetic factors would alter both intrauterine growth and adult glucose metabolism (25). The *TCF7L2* gene has consistently been associated with birth weight (26). Unfortunately, we do not have maternal genotypes available. Mother-child pair analyses suggest that the *TCF7L2* variant effect on birth weight is a reflection of its presence in the mother (26). In conclusion, genetic variants involved in fetal development, like *IGF2BP2*, may influence the response to fetal malnutrition and its consequences in the adult hypercaloric environment.

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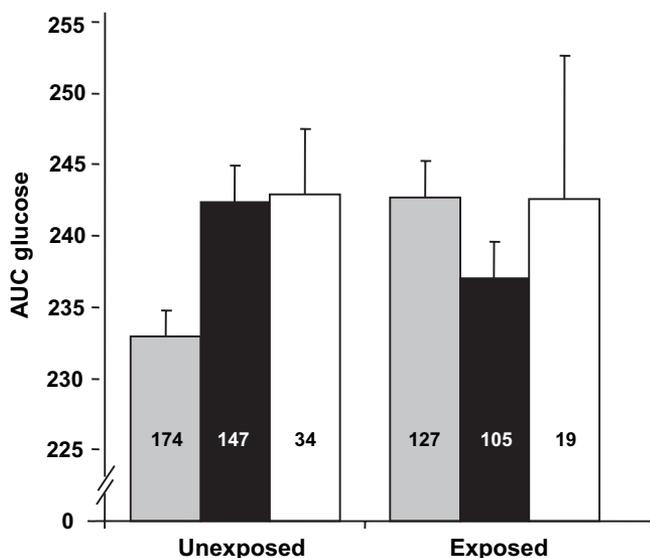


FIG. 1. The effect of the *IGF2BP2* genotypes on AUC glucose according to prenatal exposure to famine. Number of subjects shown inside of each bar, SEs of the mean shown on top of each bar. ■, GG; ■, GT; □, TT.

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