Basal and insulin mediated VLDL-triglyceride kinetics in type 2 diabetic men

Short running title: VLDL-triglyceride kinetics in type 2 diabetes

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**Objective.** Increased VLDL-TG concentration is a central feature of diabetic dyslipidemia. The objective was to compare basal and insulin mediated VLDL-TG kinetics, oxidation, and adipose tissue storage in type 2 diabetic and healthy men.

**Research Design and Methods.** Eleven type 2 diabetic and 11 healthy men, matched for BMI and age, were included. Ex vivo-labeled VLDL-TG tracers, blood- and breath samples, fat biopsies, indirect calorimetry, and body composition measures were applied to determine VLDL-TG kinetics, VLDL-TG FA oxidation, and -storage in regional adipose tissue before and during a hyperinsulinemic euglycaemic clamp.

**Results.** VLDL-TG secretion was significantly greater in diabetic compared with healthy men (Basal: 86.9 (31.0) vs. 61.9 (30.0) µmol/min, \(p=0.03\); Clamp: 60.0 (26.2) vs. 34.2 (17.9) µmol-min\(^{-1}\), \(p=0.01\)). The insulin mediated suppression of VLDL-TG secretion was significant in both groups. VLDL-TG clearance was lower in diabetic men (Basal: 84.6 (32.7) vs. 115.4 (44.3) ml-min\(^{-1}\), \(p=0.08\); Clamp: 76.3 (30.6) vs. 119.0 (50.2) ml-min\(^{-1}\), \(p=0.03\)). During hyperinsulinemia fractional VLDL-TG FA oxidation was comparable, but in percent of EE, significantly higher in diabetic men. Basal VLDL-TG storage was similar, but significantly greater in abdominal compared with leg fat.

**Conclusions.** Increased VLDL-TG in type 2 diabetic men is caused by greater VLDL-TG secretion and less so by lower VLDL-TG clearance. The ability of hyperinsulinemia to suppress VLDL-TG secretion appears preserved. During hyperinsulinemia VLDL-TG FA oxidation is significantly increased in proportion of EE in type 2 diabetic men. Greater basal abdominal VLDL-TG storage may help explain the accumulation of upper-body fat in insulin resistant individuals.

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Type 2 diabetes is associated with dyslipidemia, which is considered a major risk factor for coronary heart disease(1-3). It is characterized by hypertriglyceridemia, low high-density lipoprotein (HDL) cholesterol concentrations, small and dense low-density lipoprotein (LDL) particles, and excessive postprandial lipemia(4-7). Evidence suggests that increased concentration of very low-density lipoprotein triglycerides (VLDL-TG) is a central pathophysiological feature of diabetic dyslipidemia(4;5).

Various tracer methods have been applied to study the mechanisms responsible for hypertriglyceridemia. Most techniques are based on in vivo-labeling; i.e. infusion of labeled precursors which are eventually incorporated into either apolipoprotein B100 (apoB-100) or the TG content of the VLDL particles combined with modeling of the subsequent plasma decay data. Because only one apoB-100 molecule exists per VLDL particle, VLDL-apoB-100 reflects particle number, whereas VLDL-TG reflects the major lipid content of the particle. Studies have shown that elevated levels of VLDL-TG in type 2 diabetes is caused by a combination of increased hepatic secretion of VLDL-apoB-100(8-10) and VLDL-TG(10), specifically in the sub-fraction of large TG-rich VLDL (VLDL\(_1\))(11;12). This has focused
attention on the impact of insulin on VLDL kinetics. The acute effect of insulin on VLDL kinetics has been investigated in non-diabetic subjects (13-17). Despite different modeling approaches the results are quite unambiguous showing that acute hyperinsulinemia decreases hepatic secretion of VLDL-TG (13;14;17) and VLDL-apoB-100 (13-17), primarily VLDL₁-apoB-100 (15-17). However, to our knowledge, the effect of insulin on VLDL-TG kinetics has not been studied directly in type 2 diabetic subjects. These studies were undertaken to compare basal and insulin mediated VLDL-TG kinetics in type 2 diabetic and healthy men. A secondary objective was to assess peripheral VLDL-TG fatty acids (FA) metabolism, i.e. to what extent VLDL-TG FA are oxidized and stored in regional adipose tissue. This is relevant in the context of VLDL-TG as a potentially important energy source and its contribution to fat accumulation in specific adipose tissue depots.

**RESEARCH DESIGN AND METHODS**

The study was approved by the local ethics committee and informed consent was obtained from all participants.

**Subjects.** Eleven type 2 diabetic men and 11 healthy men with no family history of type 2 diabetes, matched for BMI and age, were recruited from the outpatient clinic and through local advertisements. All were non-smokers, weight stable, and not regularly engaged in vigorous exercise. Current diabetes treatments were lifestyle modifications alone in 6 patients and either metformin, sulfonylurea, or both in 5 patients. Oral hypoglycemic agents were discontinued three weeks prior to the study day and statins and antihypertensive drugs two weeks before. All had normal blood count, liver- and kidney function (Table 1).

**Protocol.** Screening of potentially eligible subjects was performed after an overnight 12 hour fast. A medical history was obtained, and blood samples drawn for lipid profile, HbA1c, liver and kidney function and complete blood count.

One week prior to the study day subjects meeting the eligibility criteria visited the Clinical Research Center after an overnight fast. Blood was drawn for ex vivo-labeling of VLDL-TG as described below. Dual X-ray absorptiometry (DXA) scan and abdominal CT scan at the L₂-L₃ interspace was performed to obtain anthropometric indices. Volunteers were interviewed by a dietitian, who estimated daily caloric intake, based on which, the participants consumed a weight maintaining diet (55% carbohydrate, 15% protein, 30% fat) provided by the hospital kitchen during the three days preceding the metabolic study. Volunteers were admitted to the Research Center at 22:00 the evening before the study. From this time and until the end of the study they fasted and remained in bed. At 05.30 (t=−120 min) an intravenous catheter was placed in an antecubital vein and a bolus of [9,10-³H]VLDL-TG was infused. Another catheter was placed in a dorsal hand vein for blood sampling. The hand was placed in a heated box to obtain arterialized blood. At 07.30 (t=0) 20% of the [1-¹⁴C]VLDL-TG tracer was infused as a bolus and a constant infusion with the remaining 80% was started. At t=120 min, a 5 hour infusion (1.0 mU·kg FFM⁻¹·min⁻¹) of human insulin (Actrapid; Novo Nordisk A/S) commenced. Plasma glucose was measured every 10 min and clamped at 5 mmol·l⁻¹ by variable infusion of 20% glucose. The glucose infusion rate during the last hour of the clamp (M-value) was used as an index of insulin sensitivity. Blood samples were drawn to determine VLDL-TG specific activity (SA) at t=0, 60, 80, 100 and 120 min (basal period) and 180, 240, 300, 360, 380, 400 and 420 min (clamp period). Insulin and metabolite concentrations were determined every 60 min and ApoB-100 concentration at the end of each period. Breath samples to
determine $^{14}$CO$_2$ SA were obtained at 0, 80, 100, 120, 380, 400 and 420 min. Indirect calorimetry was performed from t=60 to 80 min and from 360 to 380 min. At t=120 min fat biopsies were obtained from the abdominal (periumbilical) and femoral (inner thigh) regions using a liposuction technique. At 420 min all catheters were removed and the participants discharged. The protocol is illustrated in Figure 1.

**VLDL-TG tracer preparation.** The ex-vivo labeling procedure of VLDL-TG with radio labeled triolein has previously been described in detail(18;19). Briefly, each half of the plasma obtained from a 120 ml blood sample was mixed with either $[^1-14]$Ctri olein or $[^9,10-^3]$Htri olein and sonicated in a cell incubator at 37°C for 6 hours. The plasma was transferred to sterile Optiseal centrifuge tubes (Beckman Instruments, Inc.), covered with a saline solution ($d=1.006$ g·ml$^{-1}$) and centrifuged (Ti 50.3 rotor; Beckman Instruments, Inc.) for 18 hours at 40000 rpm and 10°C. The supernatant containing the labeled VLDL particles was removed using a sterile Pasteur pipette, filtered and stored at 5°C. Samples were tested for bacterial growth to ensure sterility.

**Body composition.** Total body fat, leg fat, fat percent, and fat free mass were examined by dual X-ray absorptiometry (DXA) scanning (QDR-2000; Hologic, Inc.). Upper-body and visceral fat mass was assessed using the CT measures of intra-abdominal and subcutaneous adipose tissue combined with abdominal fat mass measured by DXA as previously described(20). Abdominal subcutaneous fat was taken as upper body fat minus visceral fat. Leg fat was measured using the region of interest program with the DXA instrument.

**Indirect calorimetry.** Energy expenditure (EE) and substrate oxidation rates were measured by indirect calorimetry (Deltatrac monitor; Datex Instruments, Inc.) and net lipid and glucose oxidation rates were calculated by using the non-protein respiratory quotient from the above measurements(21).

**Laboratory procedures.** Plasma glucose was analyzed using an YSI 2300 STAT Plus glucose analyzer (YSI Inc.). Blood samples were placed on ice and separated as quickly as possible by centrifugation (3600 rpm at 4°C for 10 min). Aliquots of plasma (3 ml) were stored at 4°C for isolation of VLDL immediately after completion of the examination as described below. Remaining samples were stored at -20°C for later analysis. TG concentrations were analyzed using a COBAS Fara II (F. Hoffmann-La Roche Ltd). Serum insulin concentrations were measured with an immunoassay (DAKO Denmark A/S). Serum FFA concentrations were determined by a colorimetric method using a commercial kit (Wako Pure Chemical Industries, Ltd.). ApoB-100 concentration in the supernatant following ultracentrifugation (see below) was determined using an ELISA kit (Mabtech, Inc.). Duplicate samples were diluted to ensure reading on the linear part of the standard curve.

**Adipose tissue $^3$H-TG SA .** Adipose tissue lipid SA (dpm·g$^{-1}$) was measured after lipid extraction from adipose tissue biopsies as previously described(22). In brief, extracted lipid was weighed, scintillation cocktail (Optiphase HiSafe 2; PerkinElmer, Inc.) was added, mixed thoroughly, and $^3$H activity was counted.

**Breath $^{14}$CO$_2$ SA.** Breath samples were obtained using breath bags (IRIS-breath-bags; Wagner Analysen Technik). Expired air was passed through a solution containing benzethonium hydroxide (Sigma-Aldrich, Inc.) with thymolphthalein (Sigma-Aldrich, Inc.) in a scintillation vial. A color change occurred when exactly 0.25 mmol CO$_2$ was trapped in the solution. Scintillation fluid was added, and $^{14}$C activity was counted. Total CO$_2$ production rate was obtained from the indirect calorimetry measurements.
Plasma VLDL-TG concentration and SA. VLDL was isolated from approximately 3 ml of each plasma sample by ultracentrifugation as described above. The supernatant containing the VLDL fraction was obtained by tubeslicing (Beckman Instruments, Inc.) and transferred to a scintillation vial. A small sample was analyzed for TG concentration, and the plasma concentration of VLDL-TG was calculated. Scintillation fluid was added, and $^{14}$C activity was counted.

Calculations. VLDL-TG secretion rate—VLDL-TG SA steady state was effectively reached during the last hour of the basal and clamp period. VLDL-TG secretion rate ($\mu$mol·min$^{-1}$) was calculated by dividing the infusion rate by the plateau SA by the plateau SA in each period:

$$\text{VLDL-TG secretion rate} = \frac{F}{\text{SA}}$$

VLDL-TG clearance—VLDL-TG clearance rate (ml·min$^{-1}$) was calculated by dividing the secretion rate by the average VLDL-TG concentration in each period:

$$\text{VLDL-TG clearance rate} = \frac{\text{VLDL-TG secretion rate}}{C_{\text{VLDL-TG}}}$$

VLDL-TG FA oxidation—Fractional oxidation (%) of the infused [1-$^{14}$C]VLDL-TG was calculated as follows:

$$\text{Fractional VLDL-TG oxidation} = \frac{k \times \text{Ar} \times F}{\text{VLDL-TG FA oxidation rate}}$$

Here, $k$ is the volume of CO$_2$ at 20°C and 1 atm. pressure (22.4 l·mol$^{-1}$) and Ar is the fractional acetate carbon recovery factor in breath CO$_2$, and F is the tracer infusion rate. Sidossis et al. has previously calculated Ar to be 0.56 for resting conditions and 0.5 for hyperinsulinemia, respectively(23). The total VLDL-TG FA oxidation rate ($\mu$mol·min$^{-1}$) was calculated as follows and multiplied by 3 to allow for 3 FAs in each TG:

$$\text{VLDL-TG oxidation rate} = \text{Fractional VLDL-TG oxidation} \times \text{VLDL-TG oxidation rate}$$

To calculate energy production (kcal·day$^{-1}$) from VLDL-TG FA oxidation, the oxidation rate was converted to its weight equivalent using the molecular weight of oleic acid (282 g·mol$^{-1}$) and multiplied by the caloric density of 9.1 kcal·g$^{-1}$ and 1440 min·day$^{-1}$.

Adipose tissue VLDL-TG FA storage. Fractional adipose tissue VLDL-TG FA storage (%) in abdominal and leg fat was calculated using the regional (abdomen or thigh) adipose lipid SA multiplied by the total amount of lipid in the region divided by the injected dose. The VLDL-TG storage rates ($\mu$mol·min$^{-1}$) in specific regions were calculated as the fractional storage multiplied by VLDL-TG secretion rate.

Statistics. Data are mean (SD) or median (range). Between groups comparisons were performed using Student’s $t$ test or Mann-Whitney’s test. Within group comparisons were performed using Student’s $t$ test for paired comparisons or Wilcoxon’s test. Adipose tissue storage values were log transformed to obtain normal distribution before statistical processing. Correlations were tested using Pearson’s $r$ or Spearman’s rho. $p<0.05$ was considered significant.

RESULTS

Subject characteristics. Subject characteristics are summarized in Table 1. The groups were well matched for age and BMI, and there was no significant difference in body composition indices except FFM. As expected, greater HbA$_{1c}$ (6.8 (0.8) vs. 5.4 (0.4)% $p<0.001$) and TG (1.68 (1.17-3.77) vs. 1.10 (0.80-1.88) mmol·l$^{-1}$, $p<0.01$) was noted in type 2 diabetic men.

Circulating metabolites and insulin. Concentrations of glucose, TG, VLDL-TG, FFA, and insulin are shown in Figure 2. In the basal period glucose, VLDL-TG, and insulin were significantly greater in diabetic men compared with healthy men, while there was no significant difference in FFA concentrations. During the clamp, plasma glucose gradually decreased in diabetic men to the target of ~5 mmol·l$^{-1}$. While
hyperinsulinemia resulted in near-complete suppression of FFA \((p<0.001, \text{ both groups})\), the decrease in plasma VLDL-TG was more modest, but still highly significant \((p=0.001 \text{ in both groups})\). In absolute numbers the decrease in VLDL-TG was comparable \((-0.24 (0.15) \text{ vs. } -0.25 (0.11) \text{ mmol·l}^{-1}, \text{ ns})\), however, the relative decrease was significantly greater in healthy subjects \((-23.9 (18.2) \text{ vs. } -44.1 (15.7)\% \ p=0.01)\). The resulting VLDL-TG concentration was significantly greater in diabetic men compared with healthy men, whereas there was no significant difference in glucose, insulin or FFA concentrations between the groups during the clamp.

**Metabolic parameters.** Metabolic parameters are summarized in Table 2. In the basal state, glucose oxidation rates were similar but lipid oxidation rates were significantly lower in diabetic men compared with healthy men \((0.57 (0.19) \text{ vs. } 0.74 (0.14) \text{ mg·kg}^{-1}·\text{min}^{-1}, \ p=0.02)\). Conversely, there were no significant differences in glucose- and lipid oxidation rates or RQ during hyperinsulinemia. As expected, insulin-mediated glucose disposal rate was significantly reduced in diabetic men \((5.1 (3.3) \text{ vs. } 9.0 (3.7) \text{ mg·kg}^{-1}·\text{min}^{-1}, \ p=0.02)\).

**VLDL-TG secretion and clearance.** VLDL-TG SA steady state was effectively reached in the last hour of both the basal and the clamp period (Figure 3A). VLDL-TG secretion (Figure 3B) was significantly greater in diabetic men compared with healthy men in both the basal state \((86.9 (31.0) \text{ vs. } 61.9 (30.0) \mu\text{mol·min}^{-1}, \ p=0.03)\) and during hyperinsulinemia \((60.0 (26.2) \text{ vs. } 34.2 (17.9) \mu\text{mol·min}^{-1}, \ p=0.01)\). The suppression of VLDL-TG secretion was highly significant in both groups \((p<0.001, \text{ both groups})\). Moreover, the suppression was comparable between groups both in absolute numbers \((-26.9 (17.2) \text{ vs. } -27.7 (16.9) \mu\text{mol·min}^{-1}, \text{ ns})\) and in percent \((-30.6 (20.7) \text{ vs. } -43.6 (15.4)\%\, \text{ ns})\). VLDL-TG clearance (Figure 3C) was lower in diabetic men compared with healthy men in both periods although only significant only during the clamp \((84.6 (32.7) \text{ vs. } 115.4 (44.3) \text{ ml·min}^{-1}, \ p=0.08)\) and \((76.3 (30.6) \text{ vs. } 119.0 (50.2) \text{ ml·min}^{-1}, \ p=0.03)\), respectively. There was no significant change in VLDL-TG clearance in either group or in the change between the groups.

**VLDL-apoB-100 concentration and VLDL-TG/VLDL-apoB-100 ratio.** Plasma VLDL-apoB-100 concentration was significantly greater in diabetic men compared with healthy men in both the basal state \((16.3 (10.1-42.2) \text{ vs. } 12.2 (4.6-17.1) \text{ mg·dl}^{-1}, \ p=0.02)\) and during hyperinsulinemia \((12.8 (9.7-40.0) \text{ vs. } 7.7 (3.5-17.6) \text{ mg·dl}^{-1}, \ p=0.01)\). A slight, non-significant, decrease was noted in both groups during hyperinsulinemia. The decrease was not significantly different between the groups. The VLDL-TG/VLDL-apoB-100 ratio was comparable in the two groups in the basal state \((6.1 (2.0) \text{ vs. } 5.6 (1.3) \times 10^{-3} \text{ mmol·mg}^{-1}, \text{ ns})\) but significantly higher in diabetic men during hyperinsulinemia \((4.8 (1.1) \text{ vs. } 3.6 (0.9) \times 10^{-3} \text{ mmol·mg}^{-1}, \ p=0.02)\). (Figure 3E). The decrease in the ratio during the clamp was significant in both groups but less so in diabetic men, although only the relative decrease was significantly different between the groups \((-19.4 (16.9) \text{ vs. } -35.3 (14.6)\%\, \text{ ns})\).

**VLDL-TG FA oxidation.** Breath \(^{14}\text{CO}_2\) SA steady state was reached in the clamp period, but not in the basal period, allowing calculation of VLDL-TG FA oxidation during the clamp only. VLDL-TG FA oxidation during the clamp is depicted in Figure 4B-D. The fraction of VLDL-TG secretion that was oxidized was comparable in diabetic and healthy men \((44.8 (12.1) \text{ vs. } 49.6 (10.4)\%, \text{ ns})\). However, total VLDL-TG FA oxidation tended to be greater in diabetic compared with healthy men \((27.7 (14.2) \text{ vs. } 17.6 (12.2) \mu\text{mol·min}^{-1}, \ p=0.09)\). Expressed as fraction of EE, the contribution from VLDL-TG FA to total substrate oxidation was significantly
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greater in diabetic men (16.7 (7.9) vs. 9.6 (6.3)%; p=0.03).

**VLDL-TG FA adipose tissue storage.** At 120 min (4 hours after bolus infusion of [9,10-\(^3\)H]VLDL-TG), the remaining \(^3\)H plasma activity was (6.1 (2.2-20.1)% vs. 5.8 (2.3-8.0), ns, diabetic vs. healthy men, respectively). This was calculated by dividing plasma \(^3\)H activity (dpm·mL \(^{-1}\)) at 120 min by the initial activity estimated by dividing infused activity (dpm) by plasma volume (55 mL·kg FFM \(^{-1}\))(24). Fractional VLDL-TG FA storage was comparable in the groups both in abdominal (4.2 (2.9) vs. 4.2 (2.3)%, ns) and leg fat (1.0 (0.3-4.8) vs. 0.8 (0.2-5.5)%, ns) but significantly greater in abdominal compared with leg fat (Figure 5). Since practically all \(^3\)H activity had disappeared from plasma at 120 min it is possible to estimate the rate of VLDL-TG FA storage in the tissues by multiplying fractional storage by turnover rate. Despite the difference in VLDL-TG secretion rate there was no significant difference between diabetic and healthy men in fractional VLDL-TG FA oxidation during hyperinsulinemia in both type 2 diabetic and healthy men. Third, storage pattern of VLDL-TG FA is similar in both groups with significantly greater storage in abdominal subcutaneous fat compared with leg fat.

Our findings support and extend results from previous reports showing that increased postabsorptive VLDL-TG concentrations in type 2 diabetes results from hepatic hypersecretion(8-12) and adds new information regarding the effect of insulin on VLDL kinetics(13-17). In addition to increased VLDL-TG secretion we found that VLDL-TG clearance is lower in diabetic men compared with healthy men, although the difference was significant only during hyperinsulinemia. Therefore, we conclude that the increased plasma VLDL-TG concentrations observed in type 2 diabetic men is primarily explained by increased VLDL-TG secretion and only to a lesser degree by decreased VLDL-TG clearance.

To our knowledge, the effect of experimental hyperinsulinemia on VLDL-TG kinetics in type 2 diabetic patients and healthy subjects has not previously been compared directly. In one study the effect of insulin on VLDL kinetics was compared in subjects with high liver fat (80% with type 2 diabetes) and low liver fat (all non diabetic)(17). Hyperinsulinemia resulted in a rapid decline in VLDL apoB-100 secretion in the group with low liver fat, whereas there was no significant change in VLDL apoB-100 secretion in the group with high liver fat. However, the study was designed to study the effect of liver fat content rather than type 2 diabetes on VLDL kinetics. The effect of hyperinsulinemia on VLDL-apoB-100 kinetics in type 2 diabetes...
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has been studied in a few studies, but results are conflicting(25;26). However, lack of a healthy control group in the first study may explain the difference. In the present study VLDL-apoB-100 concentration was significantly greater in type 2 diabetic men compared with healthy men, and there was no change in concentrations during hyperinsulinemia. In addition, the VLDL-TG/VLDL-apoB-100 ratio, a measure of VLDL particle size, was comparable in the basal state but decreased significantly in both groups during hyperinsulinemia. This indicates that, postabsorptively, type 2 diabetic men secrete more – not larger – VLDL particles than healthy men, and that hyperinsulinemia decreases the average particle TG content without changing the particle number. This effect was, however, attenuated in type 2 diabetic men, since the relative decrease in VLDL-TG/VLDL-apoB-100 ratio was significantly greater in healthy men.

As a novel observation we found that VLDL-TG FA oxidation during hyperinsulinemia accounted for 50% of VLDL-TG turnover, equivalent to 16.7% and 9.6% of total EE, which was significantly greater in type 2 diabetic men compared with healthy men. In the basal state, the potential VLDL-TG energy yield, that is if all circulating VLDL-TG is oxidized, was ~40% of EE. Since we did not achieve steady state in breath $^{14}$CO$_2$ in the basal period we cannot estimate basal VLDL-TG FA oxidation. However, we recently reported basal VLDL-TG FA oxidation rates of ~20% of EE in obese, insulin resistant and lean women(27). In the present study the fractional oxidation of VLDL-TG FA during hyperinsulinemia was comparable in the two groups. However, due the greater VLDL-TG secretion rate, VLDL-TG oxidation rates tended to be greater in type 2 diabetic men. The greater VLDL-TG oxidation in relation to EE in type 2 diabetic men compared with healthy men suggest a decreased capacity insulin to suppress VLDL-TG oxidation in type 2 diabetic men. This is interesting in the context of “metabolic inflexibility” in type 2 diabetes, i.e. reduced capacity to shift appropriately between lipid and carbohydrate fuels for oxidation(28). Thus, we propose to extend the concept of metabolic inflexibility in type 2 diabetes to include the failure to suppress VLDL-TG FA oxidation during hyperinsulinemia.

Finally, we report that the fraction of circulating VLDL-TG FA that are stored in abdominal and leg fat is similar in type 2 diabetic and healthy men with a similar amount of fat tissue. However, the VLDL-TG FA storage rate is somewhat greater, although not significantly, in type 2 diabetic men compared with healthy men in both abdominal and leg fat (Figure 5). Both the fractional storage and the storage rate were significantly greater in abdominal fat compared with leg fat in both groups. This difference cannot be ascribed to a greater amount of abdominal fat compared with leg fat. Thus, abdominal subcutaneous fat mass was only ~25% greater than leg fat mass whereas storage was ~3 times greater in abdominal fat. Therefore, storage per kg fat was significantly greater in abdominal fat compared with leg fat in both groups. In a recent study of obese and lean women we reported similar VLDL-TG fractional storage and storage rates in abdominal and leg fat in upper body obese women. Both fractional storage and storage rates in abdominal fat were similar to the values found in men in the present study. However, leg fat storage was much less in men in the present study compared with the obese women of the previous study(27). We believe that the present study is the first to report postabsorptive VLDL-TG storage rates in regional fat in type 2 diabetic men in comparison with matched healthy men, and that the results offers new information to
explain the preferential fat accumulation in abdominal fat in upper-body obese subjects. We acknowledge the limitations of our study. The inability to detect a significant difference between groups in suppression of VLDL-TG secretion may represent a type 2 error. At least our data indicate that insulin affects VLDL-TG kinetics differently in the two groups. Basal VLDL-TG secretion rates are significantly greater in type 2 diabetic men despite significantly higher insulin concentrations. Moreover, the relative decrease in VLDL-TG concentration and VLDL-TG/VLDL-apoB-100 ratio was significantly lower in type 2 diabetic men. In addition, the capacity of insulin to reduce VLDL-TG secretion could be proportionally less pronounced at lower (physiologic) insulin levels in type 2 diabetic than in healthy men. In comparison, suppression of glucose production is impaired in type 2 diabetic subjects compared with weight-matched healthy subjects at physiologic, but not supraphysiologic, insulin concentrations(29). VLDL-TG oxidation rates were calculated by multiplying fractional oxidation by VLDL-TG turnover. Obviously, if either parameter is overestimated, the oxidation rates are overestimated. The optimal approach would be measurement of acetate recovery in each experiment, since the recovery dependents on e.g. EE and tracer infusion duration. The VLDL-TG secretion rates reported in our study are higher than values reported in most studies based on precursor labeling and compartmental modeling(30;31). However, traditional tracer dilution technique and non-compartmental modeling is a conceptually more simple approach that allows calculation of kinetic parameters of interest without the assumptions inherent to methods based on compartmental modeling. Importantly, the VLDL-TG secretion rates reported in the present study are in reasonable agreement with values from studies using other model-independent methods, e.g. splanchnic differences and studies using primed-constant reinfusion of endogenously labeled VLDL-TG tracer as discussed in detail previously(19). We also acknowledge the use of a VLDL-TG tracer prepared from VLDL-TG particles during fasting, which may not be representative to the VLDL-TG particle composition during hyperinsulinemia. Ideally, VLDL-TG sampling for tracer preparation should be performed during hyperinsulinemia. Using this approach, Lewis et al. reported unaltered VLDL-ApoB-100 kinetics(13). Moreover, we found no published information regarding the impact of altered plasma particle composition on the peripheral fate of VLDL-TG FA. Finally, we cannot extend our findings to type 2 diabetic women.

In conclusion, this study demonstrates that the increased VLDL-TG concentration in type 2 diabetic men results from increased hepatic VLDL-TG secretion, and that the ability of hyperinsulinemia to suppress VLDL-TG secretion is preserved in type 2 diabetic men at this relatively high insulin dose. Moreover, VLDL-TG FA oxidation accounts for 50% of VLDL-TG turnover, and constitutes a significantly greater proportion of EE in type 2 diabetic men (~17%) compared with age and weight matched men (10%). Finally, both type 2 diabetic and healthy men store more VLDL-TG in abdominal fat than in leg fat, which may provide a mechanism whereby some individuals develop a preferential upper-body fat distribution.

Author contributions. L.P.S. researched data, contributed to discussion, wrote manuscript, reviewed/edited manuscript. I.R.A. researched data, reviewed/edited manuscript. E.S. researched data, contributed to discussion, reviewed/edited manuscript. L.G. contributed to discussion, reviewed/edited manuscript. O.S. contributed to discussion. J.S.C. contributed to discussion, reviewed/edited manuscript. S.N. researched
data, contributed to discussion, wrote manuscript, reviewed/edited manuscript.

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Table 1: Subject characteristics

<table>
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<th>Type 2 diabetic men (n=11)</th>
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<td>53±12</td>
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<tr>
<td>Fat %</td>
<td>25.7±5.4</td>
<td>28.0±4.1</td>
</tr>
<tr>
<td>HbA₁c (%)</td>
<td>5.4±0.4</td>
<td>6.8±0.8‡</td>
</tr>
<tr>
<td>Triglycerides (mmol/l)</td>
<td>1.10 (0.80-1.88)</td>
<td>1.68 (1.17-3.77)†</td>
</tr>
<tr>
<td>Total cholesterol (mmol/l)</td>
<td>4.6±0.7</td>
<td>4.9±0.6</td>
</tr>
<tr>
<td>LDL-cholesterol (mmol/l)</td>
<td>3.0±0.6</td>
<td>3.0±0.5</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/l)</td>
<td>1.1 (0.8-1.5)</td>
<td>0.9 (0.7-1.3)</td>
</tr>
</tbody>
</table>

Data are mean ± SD or median (range). BMI = body mass index; FFM = fat free mass; sc = subcutaneous. *p<0.05, †p<0.01, ‡p<0.001 vs. healthy men

Table 2. Circulating metabolites, insulin, and metabolic parameters in the basal state and during hyperinsulinemia.

<table>
<thead>
<tr>
<th></th>
<th>Healthy men (n=11)</th>
<th>Type 2 diabetic men (n=11)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Basal</td>
<td>Clamp</td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>5.5 (4.5-6.7)</td>
<td>4.9 (4.6-5.3)</td>
</tr>
<tr>
<td>FFA (mmol/l)</td>
<td>0.43±0.12</td>
<td>0.03 (0.02-0.16)</td>
</tr>
<tr>
<td>VLDL-TG (mmol/l)</td>
<td>0.54 (0.31-1.10)</td>
<td>0.28 (0.13-0.63)</td>
</tr>
<tr>
<td>Insulin (pmol/l)</td>
<td>27 (18-66)</td>
<td>320±31</td>
</tr>
<tr>
<td>EE (kcal/day)</td>
<td>1956±125</td>
<td>2021±131</td>
</tr>
<tr>
<td>RQ</td>
<td>0.81±0.04</td>
<td>0.89±0.05</td>
</tr>
<tr>
<td>Glucose oxidation (mg/kg/min)</td>
<td>1.02±0.52</td>
<td>2.09±0.72</td>
</tr>
<tr>
<td>Lipid oxidation (mg/kg/min)</td>
<td>0.74±0.14</td>
<td>0.37±0.23</td>
</tr>
<tr>
<td>GIR (mg/kg/min)</td>
<td>9.0±3.7</td>
<td>5.1±3.3*</td>
</tr>
</tbody>
</table>

Data are mean ± SD or median (range). FFA = free fatty acids; VLDL-TG = very-low-density-lipoprotein triglyceride; EE = energy expenditure; RQ = respiratory quotient; GIR = glucose infusion rate. *p<0.05, †p<0.01, ‡p<0.001 vs. healthy men. §p<0.05, ||p<0.01 difference (basal-clamp) vs. difference (basal-clamp) in healthy men.
Figure legends
Figure 1. Study protocol.
Figure 2. Glucose infusion rate during the hyperinsulinemic clamp (A), concentrations of glucose (B), TG (C), VLDL-TG (D), FFA (E), and insulin (F) in the basal state and during the hyperinsulinemic clamp. * p<0.05 between groups (60-120 min and 360-420 min). Closed circles, healthy subjects; open circles, type 2 diabetic subjects. Data are presented as mean±SEM.
Figure 3. VLDL-TG specific activity (A), VLDL-TG secretion rate (B), VLDL-TG clearance rate (C), VLDL-TG concentration (D), and VLDL-TG / VLDL-ApoB-100 ratio (E) in the VLDL-TG SA steady state periods. Closed circles and bars, healthy subjects, open circles and bars, type 2 diabetic subjects. Data are mean±SEM.
Figure 4. Breath $^{14}$CO$_2$ SA steady state was reached in the clamp period, but not in the basal period. Therefore, only VLDL-TG FA oxidation data from the clamp period are illustrated and analyzed statistically. VLDL-TG FA oxidation expressed as fraction of secretion (B) and as oxidation rate (fractional oxidation x VLDL-TG secretion rate) (C). VLDL-TG FA oxidation as fraction of EE (D). Closed circles and bars, healthy subjects; open circles and bars, type 2 diabetic subjects. Data are mean±SEM.
Figure 5. Storage of VLDL-TG FA in abdominal (A-B) and leg (C-D) subcutaneous adipose tissue expressed as fraction of VLDL-TG secretion (A and C) and as storage rate (fraction storage x VLDL-TG secretion rate) (B and D). Closed bars, healthy subjects; open bars, type 2 diabetic subjects. Data are presented as mean±SEM.

Figure 1
Figure 2

- **Healthy subjects**
- **Type 2 diabetic subjects**

**A**
- Basal vs. Clamp for GIR (mg/kg FF/min).

**B**
- Basal vs. Clamp for Plasma glucose (mmol/l).

**C**
- Basal vs. Clamp for Plasma TG (mmol/l).

**D**
- Basal vs. Clamp for Plasma VLD-TG (mmol/l).

**E**
- Basal vs. Clamp for Serum FFA (mmol/l).

**F**
- Basal vs. Clamp for Serum Insulin (pmol/l).
Figure 3

A

B

C

D

E

VLDL-triglyceride kinetics in type 2 diabetes